

# Maximisation of Peak Capacity for Peptide Mapping using a Column Coupling Approach with Solid-Core Columns

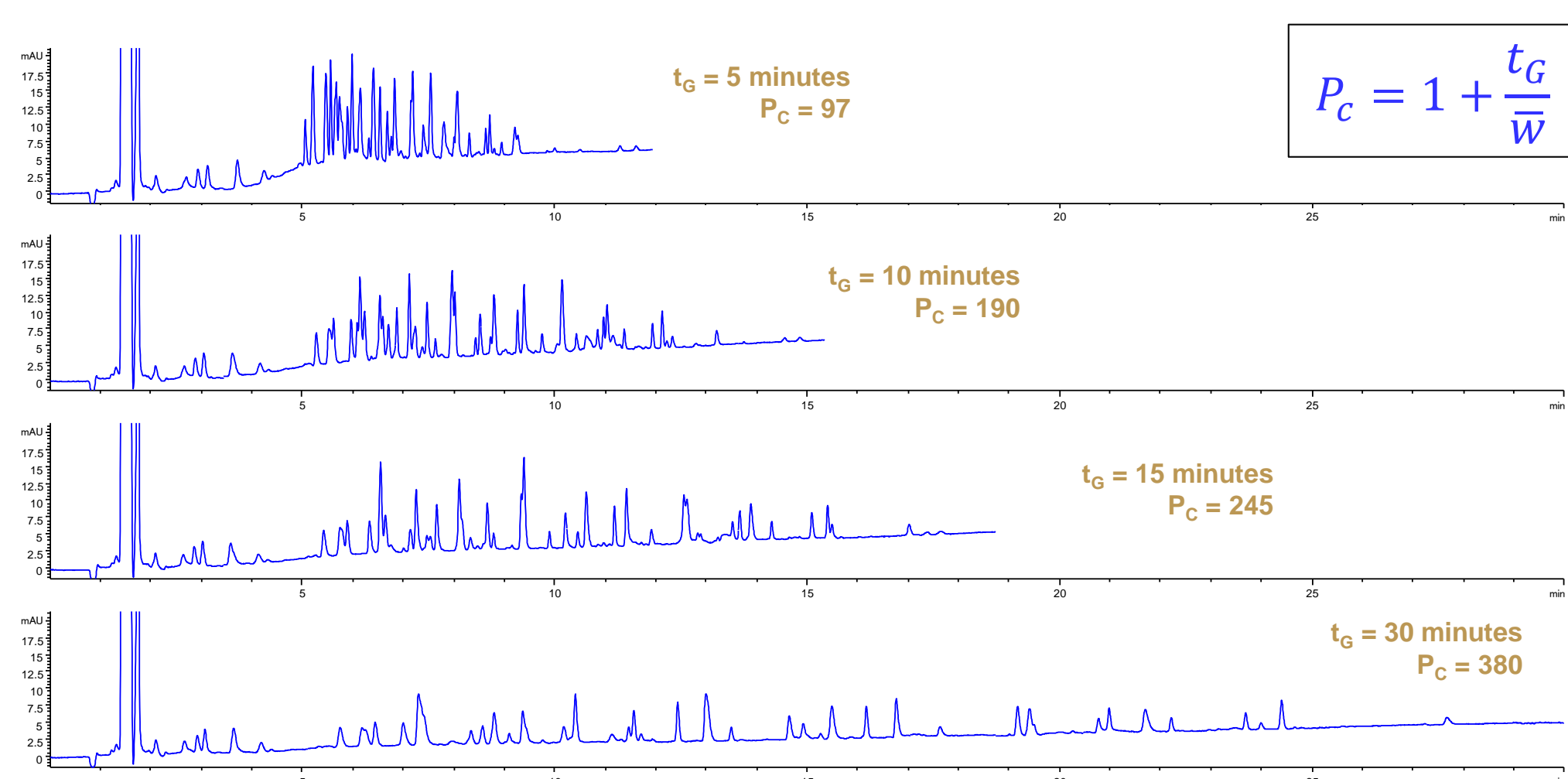
Alan P McKeown

Advanced Chromatography Technologies Ltd, 1 Berry St, Aberdeen, Scotland, AB25 1HF UK

## 1. Introduction

- Peptide mapping is used to monitor changes in the primary amino acid sequence of a protein.
- Changes to the amino acid sequence are known as Critical Quality Attributes as they can greatly affect the safety and efficacy of a biopharmaceutical drug.
- Proteins are enzymatically digested to produce characteristic peptide fragments which can then be analysed using HPLC.
- Peak capacity ( $P_C$ ) is particularly important in peptide mapping due to the large number of peaks present.
- This poster presents work performed to optimise the peak capacity for peptide mapping applications on a high performing solid-core column (**ACE UltraCore 2.5 SuperC18**).
- Peak capacity was further enhanced by coupling multiple columns in series.

## 3. Gradient Time Optimisation

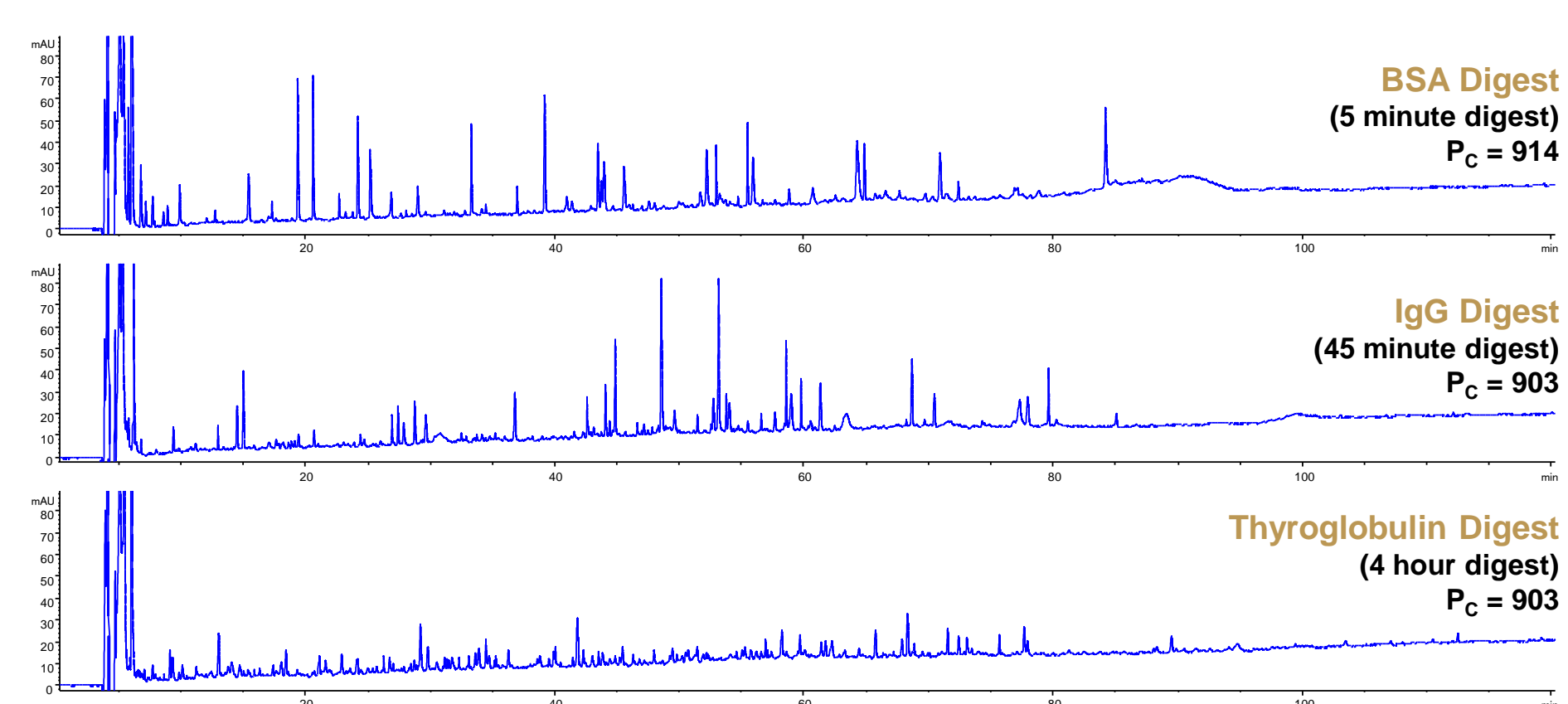


- 10 – 40 %B gradient, 0.21 mL/min.
- Pre-digested BSA sample.
- Highest peak capacity seen with longest gradient.

## 5. Column Coupling

- Complex samples such as tryptic digests can benefit from the improved resolution achieved by increasing column length.
- The high pressure limits of UHPLC instruments (>1,000 bar) allow multiple columns to be connected in series to increase effective column length.
- **ACE UltraCore 2.5**  $\mu$ m solid-core particles provide performance similar to 1.7  $\mu$ m fully porous particles at much lower back pressure – ideal for connecting multiple columns.
- The optimised single column method (150 x 2.1 mm) was translated to 3 coupled columns totalling 450 mm in length.
- The final method was then applied to the analysis of three freshly prepared protein digest samples.

## 7. Application to other Protein Digests Samples



### Method Conditions

Column: **ACE UltraCore 2.5 SuperC18** (450 x 2.1 mm total)  
 Mobile phases: A: 0.05 % TFA in H<sub>2</sub>O  
 B: 0.05 % TFA in MeCN  
 Gradient: 10 – 40 % B in 90 mins then 40 – 65 % B in 30 mins  
 Temperature: 60 °C  
 Flow rate: 0.21 mL/min  
 Injection volume: 20  $\mu$ L  
 Detection: UV, 214 nm

### Samples

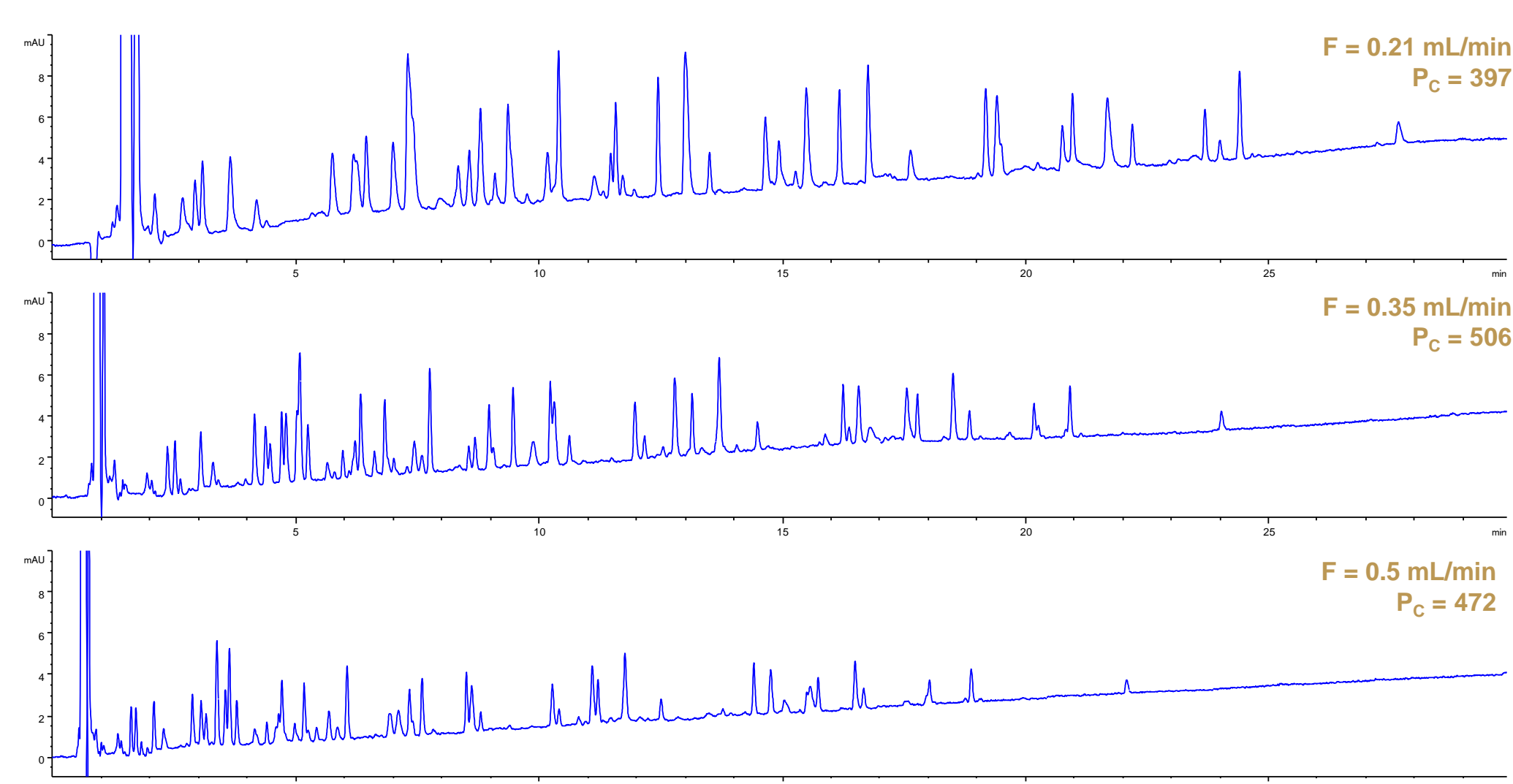
➤ BSA (66.5 kDa), IgG (150 kDa) and thyroglobulin (660 kDa) proteins were digested in this project.  
 ➤ Native proteins were digested using Thermo SMART Digest kit. (uses a combination of immobilised trypsin enzyme and heat to digest proteins).  
 ➤ Digestion time determined from kit protocols.  
 ➤ Digestions were very reproducible.

## 2. Method Optimisation

- Gradient time and flow rate were optimised using a pre-digested BSA sample.
  - $t_G$  = 5, 10, 15, 30 minutes
  - F = 0.21, 0.35, 0.5 mL/min
- An **ACE UltraCore 2.5 SuperC18** solid-core column was used for method optimisation.
- Increased efficiency of superficially porous stationary phase improved resolution of closely related peaks.

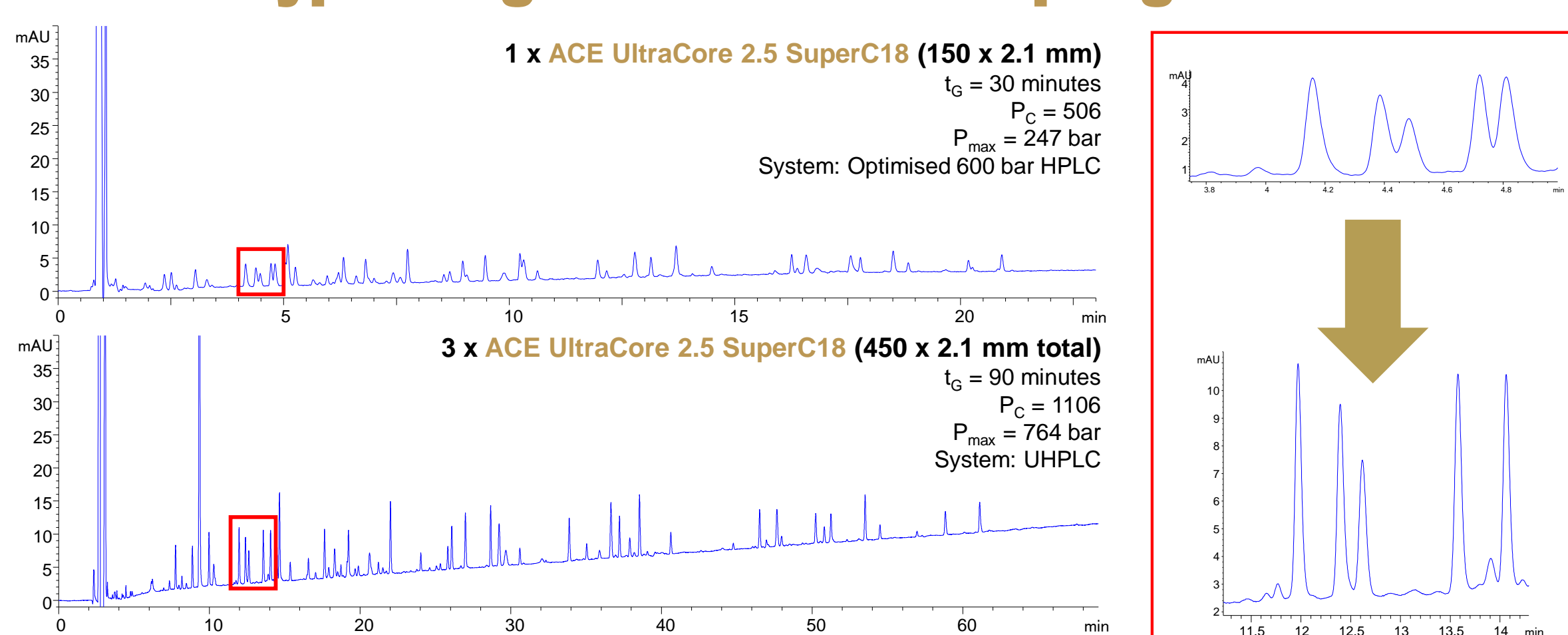
Column:	ACE UltraCore 2.5 SuperC18, 150 x 2.1 mm
Part number:	CORE-25A-1502U
Mobile phases:	A: 0.05 % TFA in H <sub>2</sub> O B: 0.05 % TFA in MeCN
Injection volume:	20 $\mu$ L
Temperature:	60 °C
Detection:	UV, 214 nm

## 4. Flow Rate Optimisation



- 10 – 40 %B gradient in 30 mins.
- Highest peak capacity observed at 0.35 mL/min.

## 6. BSA Tryptic Digest - Column Coupling



- Large increase in peak capacity with coupled columns, 3-fold increase in back pressure.
- Additional sample complexity revealed.

### Method Conditions

Mobile phases: A: 0.05 % TFA in H<sub>2</sub>O  
 B: 0.05 % TFA in MeCN  
 Gradient: 10 – 40 % B  
 Temperature: 60 °C  
 Flow rate: 0.35 mL/min  
 Injection volume: 20  $\mu$ L  
 Detection: UV, 214 nm

## 8. Summary and Conclusions

- A generally applicable, optimised gradient method was produced for the high resolution analysis of tryptic digest samples.
- Peak capacity of 506 achieved with a single **ACE UltraCore 2.5 SuperC18** column using optimised method parameters.
- Three **ACE UltraCore 2.5 SuperC18** columns were coupled together to increase peak capacity to 1106.
- High efficiency of **ACE UltraCore** particles increased the resolution of the separation.
- A Thermo SMART digest kit was used to prepare three different digests that were successfully analysed using the column coupling approach.